

Außervillgraten, 26 June 2015
Haus Valgrata



ÖGTPM
Österreichische Gesellschaft für
Tropenmedizin, Parasitologie und
Migrationsmedizin



PARASITOLOGISCHE FACHGESPRÄCHE 2015
PARASITOLOGICAL DAY 2015

“VEKTOREN & ERREGER”
“VECTORS & AGENTS”



Programme/Abstracts¹

Publisher: Österreichische Gesellschaft für Tropenmedizin, Parasitologie und Migrationsmedizin,
Kinderspitalgasse 15, 1090 Wien; Wien, Juni 2015
Editors: Georg Duscher, Christoph Hörweg, Gernot Walder
Print: Naturhistorisches Museum Wien

¹ Abstracts in chronological order



Wann/When **Fr 26. Juni 2015 / June 26, 2015**
Wo/Where Kultursaal Außervillgraten, Haus Valgrata, A-9931 Außervillgraten 136b

- 09.30 – 09.40 **WELCOME ADDRESS**
Josef MAIR (Mayor of the village of Außervillgraten), Gernot Walder (Local organizer)
- 09.40 – 11.00 **MOSQUITOES & CO.** (Chair: Hans-Peter FÜHRER)
- 09.40 – 10.00 **Julia BUTTER**, Sarah Übleis, Dominik Berer, Barbara Eigner, Carina Zित्रa, Karin Lebl, Hans-Peter Führer: Filaroid helminths and avian malaria in mosquitoes in metropolitan Vienna in 2014
- 10.00 – 10.20 **Michael KOTHMAYER**, Eva Flechl, Carina Zित्रa, Karin Lebl, Adelheid Obwaller, Hans-Peter Führer: An overview on the *Culex pipiens*-complex in metropolitan Vienna
- 10.20 – 10.40 **Hans-Peter FÜHRER**: Mosquitoes and Mosquito Borne Parasitic Diseases in Eastern Austria
- 10.40 – 11.00 **Gernot WALDER**, Josef Simeoni, Alberta Stenico: The advance of the tiger mosquito in the Tyrols 2009-2014.
- 11.00 – 11.30 *Coffee break*
- 11.30 – 12.15 **DIGENEANS & WATER SNAILS** (Chair: Christoph HÖRWEG)
- 11.30 – 11.50 **Christian HUSCH**, Helmut Sattmann, Julia Walochnik: Diversity of *Fascioloides magna* and *Fasciola hepatica* in Austria
- 11.50 – 12.10 **Nadine HOHENSEE**, Julia Walochnik, Christoph Hörweg, Helmut Sattmann: Digenean trematodes in freshwater snails in the surroundings of Vienna with a focus on species involved in human infections
- 12.15 – 13.15 *Lunch break*
- 13.15 – 15.45 **PARASITES OF DEER & FOXES** (Chair: Georg DUSCHER)
- 13.15 – 13.35 **Naike MROWIETZ**, Rita Cezanne, Verena E. Habler, Barbara Eigner, Adnan Hodzic, Georg G. Duscher, Judit Lazar, Walter Glawischnig, Hans-Peter Führer: Blood parasites of red foxes in Tyrol and Vorarlberg (2014/15)
- 13.35 – 13.55 **Adnan HODZIC**, Amer Alić, Hans-Peter Führer, Josef Harl, Walpurga Wille-Piazzai, Georg G. Duscher: Study on vector-borne pathogens in red foxes from Bosnia and Herzegovina
- 13.55 – 14.15 **Natascha LEITNER**, Laurin Schwarzmann, Carina Zित्रa, Nicola Palmieri, Barbara Eigner, Domenico Otranto, Walter Glawischnig, Hans-Peter Führer: Larvae causing nasopharyngeal myiasis of red deer (*Cervus elaphus*) from Vorarlberg and Tyrol – morphological and molecular assessment
- 14.15 – 14.35 **Georg G. DUSCHER**, Monika Wetscher, Raphaela Baumgartner, Gernot Walder: Roe deer serum surveillance for the occurrence of antibody against TBE: additional data for risk assessment in Austria
- 14.35 – 14.55 **Josef SIMEONI**: Zecken und durch sie übertragbare Infektionskrankheiten in Südtirol
- 14.55 – 15.05 **FINAL DISCUSSION / CLOSING WORDS**
- 15.15 – 16.15 Guided tour through the **Labor für Infektionsdiagnostik Dr. Gernot Walder GmbH**, A-9931 Außervillgraten 30

Filarioid helminths and avian malaria in mosquitoes in metropolitan Vienna in 2014

Julia Butter¹, Sarah Übleis¹, Dominik Berer², Barbara Eigner¹, Carina Zित्रा¹, Karin Lebl², Hans-Peter Fuehrer¹

¹Institute of Parasitology, Department of Pathobiology, University of Veterinary Medicine Vienna, Austria

²Institute for Veterinary Public Health, University of Veterinary Medicine Vienna, Austria

e-mail: hans-peter.fuehrer@vetmeduni.ac.at

A range of factors (e.g. climate change, trade and people/pet movement) have major impacts on the distribution of invertebrate vectors (e.g. mosquitoes), invasive pathogens (e.g. *Dirofilaria repens*) and their vertebrate hosts (humans, pets, livestock and wild animals). This is leading to an emergence and spread of vector-borne infections. First findings of *D. repens* in mosquitoes in Burgenland in 2012 indicate that this parasite invaded Austria. However, there is virtually no information about the parasite fauna transmitted by mosquitoes in metropolitan Vienna.

Within this study more than 8,500 mosquitoes were caught in 2014 at three different sampling sites in Vienna (e.g. at the Lobau area), using specialized traps equipped with carbon dioxide. Mosquitoes were specified according to morphological characteristics, pooled and homogenized. After DNA extraction each pool was screened for the presence of DNA of filarioid helminths and *Plasmodium* spp. using molecular tools.

DNA of several vector borne parasitic pathogens was examined within this study – e.g. avian malaria. Furthermore the results of the 2013 surveillance are compared with the results of 2014.

Neither *D. repens* nor *D. immitis* were found in mosquitoes in metropolitan Vienna - indicating that these parasites were not present in the examined area during the investigation period in 2013 and 2014.

This study is supported by the Austrian Science Fund (FWF): I1437

An overview on the *Culex pipiens*-complex in metropolitan Vienna

Michael Kothmayer¹, Eva Flechl¹, Carina Zित्रा¹, Karin Lebl², Adelheid Obwaller³, Hans-Peter Fuehrer¹

¹Institute of Parasitology, Department of Pathobiology, University of Veterinary Medicine Vienna, Austria

²Institute for Veterinary Public Health, University of Veterinary Medicine Vienna, Austria

³Federal Ministry of Defence and Sports, Division of Science, Research and Development, Vienna, Austria

e-mail: hans-peter.fuehrer@vetmeduni.ac.at

The *Culex pipiens*-complex consists of several subtypes, species and forms like *Cx. pipiens pipiens*, *Cx. pipiens molestus* and *Cx. torrentium*. Each form has its own ecological and host preferences and may act as vector of different pathogens. However, morphological discrimination is not possible in most cases.

In our study 2 permanent sampling sites in Vienna were monitored using specialized mosquito traps equipped with carbon dioxide as attractant. Female mosquitoes were sampled twice a month from April to October 2014. Mosquitoes were specified to species/complex level and stored at -80°C until further analysis. Overall DNA of 313 mosquitoes out of the *Cx. pipiens*-complex was extracted and analyzed with molecular tools allowing genotyping of this complex.

Primary results indicate that various forms of this complex are present in metropolitan Vienna and further investigations are of urgent need to prove and examine the vector competence for various diseases of those forms.

This study is supported by the Austrian Science Fund (FWF): I1437

Mosquitoes and Mosquito Borne Parasitic Diseases in Eastern Austria

Hans-Peter Fuehrer

Institute of Parasitology, Department of Pathobiology, University of Veterinary Medicine Vienna, Austria
e-mail: hans-peter.fuehrer@vetmeduni.ac.at

More than 40 mosquito species have been reported in Austria. Of those four species are neobiota, namely *Aedes albopictus* (Asian tiger mosquito), *Ochlerotatus japonicus japonicus* (Asian bush mosquito), *Culiseta longiareolata* and *Anopheles hyrcanus*. Several species of the Austrian mosquito fauna are able to act as vectors of various Mosquito Borne Diseases (e.g. *Dirofilaria* spp.).

An overview on the distribution of mosquitoes and Mosquito Borne Parasitic Diseases in Eastern Austria will be given. Moreover, novel diagnostic tools (e.g. Direct PCR) for the molecular analysis of vectors and pathogens will be presented.

This study is supported by the Austrian Science Fund (FWF): I1437

The advance of the tiger mosquito in the Tyrols 2009-2014

Gernot Walder¹, Josef Simeoni², Alberta Stenico²

¹ Labor für Infektionsdiagnostik Dr. Gernot Walder GmbH, 9931 Außervillgraten 30, Austria

² Dienst für Hygiene und öffentliche Gesundheit des Gesundheitsbezirkes Bozen, Amba Alagistr. 33, 39100 Bozen, Italien
e-mail: gernot.walder@i-med.ac.at

In 2009 tiger mosquitoes were first detected in South Tyrol in the city of Bozen. In the following years, the abundance of the emerging vector was monitored by ovi-traps between Salurn and Innsbruck as well as between Meran and Lienz. In 2010 increasing numbers of mosquitoes were recorded in Bozen, in 2011 also neighbouring communities were affected. Until 2014 the vector has spread as far as Sterzing, Meran and eventually Bruneck. The total number of collected eggs increased every year and a further geographic spread is expected over the next years. Up to now, no eggs were recorded north or east of the local continental divide.

Diversity of *Fascioloides magna* and *Fasciola hepatica* in Austria

Christian Husch¹, Helmut Sattmann², Julia Walochnik¹

¹ Institut für Spezifische Prophylaxe und Tropenmedizin, Kinderspitalgasse 15, A-1090 Wien

² Naturhistorisches Museum Wien, 3. Zoologische Abteilung, Burgring 7, A-1010 Wien

e-mail: helmut.sattmann@nhm-wien.ac.at

Fascioloides magna was introduced into Europe in the 2nd half of the 19th century with game animals from North America and mainly infects the liver of deer. Roe deer, sheep and goats are dead-end hosts and usually die within 6 months after infection. Humans are not known to be a host.

The common liver fluke *Fasciola hepatica* is a parasitic flatworm which occurs in over 50 countries and infects the liver of various mammals including humans. It is estimated that around 2.4 million humans worldwide are infected with *F. hepatica*. As the main hosts are sheep and cattle *F. hepatica* also has a great impact on meat production and leads to significant economy loss annually.

The aim of this study was to investigate the genetic diversity of *F. magna* and *F. hepatica* in Austria. Altogether, 26 individuals of *F. magna* isolated from deer from the floodplains south of Vienna, the only known endemic region in Austria, and 33 individuals of *F. hepatica*, obtained from cattle from various regions in Austria, were analysed by molecular biological methods. To distinguish the different haplotypes the sequences of the conserved mitochondrial genes COX1 and NAD1 were examined and compared with reference strains. It was shown that all 26 individuals of *F. magna* investigated are of the same haplotypes, namely the haplotypes COX1-Ha3 and NAD1-Ha4, respectively. These are common also in Hungary, Slovakia, the Czech Republic and Croatia.

The 33 individuals of *F. hepatica* investigated could be grouped into 4 different clusters, whereby the most common genotype was shown by 22 individuals and the second most common by 8 individuals. Two genotypes were represented only by one individual each. Differences were more pronounced in the NAD1 gene than in the COX1 gene.

Altogether, our data indicate that the Austrian *F. magna* population is rather homogenous and might even be the result of a single introduction into Austria, while the Austrian *F. hepatica* population is, as expected and as also known from other countries, rather diverse.

Digenean trematodes in freshwater snails in the surroundings of Vienna with a focus on species involved in human infections

Nadine Hohensee^{1,2}, Julia Walochnik¹, Christoph Hörweg², Helmut Sattmann²

¹ Institute of Specific Prophylaxis und Tropical Medicine, Medical University of Vienna, Kinderspitalgasse 15, 1090 Vienna, Austria

² Natural History Museum Vienna, Burgring 7, 1010 Vienna, Austria
e-mail: nadine@hohensee.de

Cercariae of different species of bird schistosomes are known to be the causative agents of human cercarial dermatitis, generally known as swimmer's itch. Besides *Trichobilharzia* species, which are the most frequent cause for this disease, infections may also be caused by related genera, including *Gigantobilharzia*, *Bilharziella* and *Austrobilharzia*. The most important final hosts are waterfowl of the family Anatidae, however, humans may be infested when the cercariae accidentally penetrate the skin of bathing humans. In Europe, several foci with high prevalence are known, including regions in Austria. Due to proceeding anthropogenic changes in the environment and increasing eutrophication, the distribution of bird schistosomes is expanding and cercarial dermatitis has been recognized as an emerging disease in Europe.

In contrast, echinostomiasis is a foodborne disease. At least 20 species of 8 genera are known to cause human echinostomiasis, of which the genus *Echinostoma* is the most important. The metacercariae encyst in freshwater snails, fish, mollusks, crustaceans and frogs and are taken up orally by the final host. The disease is endemic to Southeast Asia due to food patterns, however, the pathogens occur worldwide and there is a potential risk of infection all over the world.

Detailed knowledge on the prevalence of medically important digeneans in Austria is important for a profound risk assessment. Furthermore, some digeneans are also of veterinary importance. In the current project we investigate several locations in the surroundings of Vienna and collect the main intermediate hosts, *Lymnaea stagnalis* and *Radix labiata*. We collected 380 individuals of these species in July 2014, and collecting will be continued in summer 2015. Until now, 169 individuals of *R. labiata* and 6 of *L. stagnalis* were infected with digeneans, the majority being echinostomatid species. No schistosomatid species were found yet. An overview of the digeneans found will be presented.

Blood Parasites of Red Foxes in Tyrol and Vorarlberg (2014/15)

Naike Mrowietz¹, Rita Cezanne¹, Verena Elisabeth Habler¹, Barbara Eigner¹, Adnan Hodzic¹, Georg G. Duscher¹, Judit Lazar², Walter Glawischnig², Hans-Peter Fuehrer¹

¹Institute of Parasitology, Department of Pathobiology, University of Veterinary Medicine Vienna, Austria

²Institute for Veterinary Disease Control Innsbruck, Austrian Agency for Health and Food Safety, Austria

e-mail: hans-peter.fuehrer@vetmeduni.ac.at

The red fox (*Vulpes vulpes*) has been described as carrier of many human and pet relevant pathogens such as rabies, fox tapeworms and scabies. In the recent past it was also assumed in the spreading and transmission of several tick-borne parasitic pathogens like *Hepatozoon canis* and *Babesia vulpes*. Previously a molecular screening for several tick-borne pathogens of spleens and blood originating from 36 foxes from Eastern Austria revealed a high amount of *Babesia microti*-like and *Hepatozoon canis* positive animals.

Within this study 353 blood samples of foxes from Tyrol and Vorarlberg were sampled on filter paper in 2014/2015 and analysed for (zoonotic) vector borne diseases with molecular tools. First results indicate that several pathogens like *Hepatozoon canis* and *Babesia vulpes* (formerly *B. microti*-like) are present in the examined areas.

It can be concluded that foxes are carriers of several pathogens (of veterinary and/or medical importance) in Tyrol and Vorarlberg.

Study on vector-borne pathogens in red foxes from Bosnia and Herzegovina

Adnan Hodžić¹, Amer Alić², Hans-Peter Fuehrer¹, Josef Harl¹, Walpurga Wille-Piazzai¹, Georg G. Duscher¹

¹ Institute of Parasitology, Department of Pathobiology, University of Veterinary Medicine Vienna, Vienna, Austria

² Department of Pathology, Veterinary Faculty, University of Sarajevo, Sarajevo, Bosnia and Herzegovina
e-mail: adnan.hodzic@vetmeduni.ac.at

The aim of this study was to investigate the occurrence and geographical distribution of *Babesia* spp., *Hepatozoon canis*, *Anaplasma* spp., *Bartonella* spp., 'Candidatus Neoehrlichia mikurensis', *Ehrlichia canis*, *Rickettsia* spp. and blood filaroid nematodes in red foxes from Bosnia and Herzegovina.

Spleen samples were collected from total of 119 red foxes originated from six geographical regions and examined for the presence of several vector-borne pathogens by conventional PCRs and sequencing. Three species of apicomplexan parasites were identified by molecular methods in 73 (60.8%) animals. *B. canis*, *B. cf. microti* and *H. canis* were detected in 1 (0.8%), 38 (31.9%) and 46 (38.6%) spleen samples, respectively. No statistically significant differences between geographical region, sex or age of the host in the infection prevalence of *B. cf. microti* were observed, but females (52.9%; 18/34) were significantly more infected with *H. canis* than males (32.9%; 28/85).

The presence of DNA of vector-borne bacteria and blood filaroid nematodes could not be confirmed in our study. This study reports, for the first time, the occurrence of *B. canis*, *B. cf. microti* and *H. canis* parasites in foxes from Bosnia and Herzegovina. Moreover, the relatively high prevalence of *B. cf. microti* and *H. canis* support the existence of a sylvatic cycle and reinforces the assumption that this wild canid species might be a possible reservoir and source of infection for domestic dogs.

Larvae causing nasopharyngeal myiasis of red deer (*Cervus elaphus*) from Vorarlberg and Tyrol – morphological and molecular assessment

Natascha Leitner¹, Laurin Schwarzmann¹, Carina Zittra¹, Nicola Palmieri¹, Barbara Eigner¹, Domenico Otranto², Walter Glawischnig³, Hans-Peter Fuehrer¹

¹Institute of Parasitology, Department of Pathobiology, University of Veterinary Medicine Vienna, Austria

²Dipartimento di Medicina Veterinaria, Università degli Studi di Bari, Bari, Valenzano, Italy

³Institute for Veterinary Disease Control Innsbruck, Austrian Agency for Health and Food Safety, Austria

e-mail: hans-peter.fuehrer@vetmeduni.ac.at

Nasopharyngeal myiasis are caused by larvae of bot flies (Diptera, Oestridae, Oestrinae), which have evolved a high host specificity for the hosts they parasitize. In 2014 bot flies (n= 917) were collected in 138 (58.2%) out of 237 red deer (*Cervus elaphus* Linnaeus, 1758) hunted in Vorarlberg and Tyrol (Austria). After being stored in 70% ethanol larvae were identified to species level and developmental stage using morphological and morphometric keys (e.g., measurements of body length, head capsule width). Larvae were also molecularly characterized by PCR amplification and partial sequence analysis of the mitochondrial cytochrome oxidase 1 gene. Morphological and genetic data were concordant in identifying larvae as belonging to *Cephenemyia auribarbis* and *Pharyngomyia picta*. In addition, genetic variations were also found within the specimens collected in both geographical locations.

Roe deer serum surveillance for the occurrence of antibody against TBE: additional data for risk assessment in Austria

Georg G. Duscher¹, Monika Wetscher², Raphaela Baumgartner², Gernot Walder²

¹Institute of Parasitology, Department for Pathobiology, University of Veterinary Medicine Vienna, Vienna, Austria

²Dr. Gernot Walder GmbH, Außervillgraten, Austria

e-mail: georg.duscher@vetmeduni.ac.at

Tick-borne encephalitis (TBE) is one of the serious pathogens transmitted by ticks, especially by *Ixodes ricinus*. Due to a long history of scientific efforts and public information, the Austrian citizens are aware of the tick-transmitted virus. So, 85% of the inhabitants of Austria are vaccinated at least once in their life against TBE virus. Anyhow, it is still important to search for the occurrence and distribution of new infection foci of TBE.

One possibility to look for the virus is to investigate ticks, which is rather time consuming and acquire high costs due to the high number of ticks required to obtain reliable data. Another possibility is to analyse the recorded human cases. There are maps drawn based on these cases, which might be biased due to the high vaccination rates, migration and traveling habits of the humans. Further strategies to define infection foci are to use wildlife animals as indicator species to detect antibodies in the serum. Many scientists looked in the rodents for such antibodies. This setup can deliver very good results, but in terms of large scale surveillances this also is time and cost consuming. So, we have chosen roe deer sera to investigate for the occurrence of antibodies against TBE. This animal species has a small home range and can be found all over the country. Sampling is rather easy, because roe deer is shot regularly to keep the population on a certain ecological level.

We obtained 945 sera originating from all over Austria. Further on we screened the samples with IFAT. Twenty-two positive and 17 samples with a borderline titre were found. About 40% of the positive samples and 70% of the borderline titre sera were found in areas which are not overlapping with areas of TBE occurrence in humans. Further efforts are needed to confirm or reject the new endemic foci by investigating ticks, rodents and roe deer.

Zecken und durch sie übertragbare Infektionskrankheiten in Südtirol

Josef Simeoni

Dienst für Hygiene und öffentliche Gesundheit des Gesundheitsbezirkes Bozen, Amba Alagistr, 33, 39100 Bozen, Italien
e-mail: josef.simeoni@sabes.it

Die aktive und systemische Überwachung der Zecken und der durch sie übertragbaren Infektionskrankheiten begann in Südtirol Mitte der 80er Jahre.

Vorkommen der Zecken

Bei verschiedenen südtirolweiten Zeckensammlungen konnte eine Mappe über die Verbreitung der Zecken angelegt werden.

I. ricinus: Großraum Bozen, Überetsch, Unterland, Etschtal und Meran-Burggrafenamt u. Vinschgau, in geringerem Ausmaß im Eisacktal bis Sterzing und sporadisch im Pustertal - Eingang Ahrntal;

D. marginatus u. *H. punctata*: im Vinschgau

I. hexagonus u. *R. sanguineus*: sporadisch im Raum Unterland.

Lyme-Borreliose

Die ersten klinischen Fälle wurden Mitte der 80er Jahre im Überetsch und Großraum Bozen diagnostiziert. In den darauffolgenden Jahren wurden südtirolweit konstant 5 bis 15 Fälle von Borreliosen gemeldet mit Schwerpunkt Überetsch u. Unterland. Serologische Studien ergaben (fallende Inzidenz): *B. afzelii*, *B. garinii*, *B. burgdorferi* s.s. u. *B. valaisiana*.

FSME

Der erste klinische Fall von FSME konnte im Jahr 2000 im Überetsch beobachtet werden, in den darauffolgenden Jahren wurden regelmäßig 1-2 FSME-Fälle im Überetsch und Unterland gemeldet. 2006 konnte der erste kulturelle Nachweis eines FSME-Virus aus *I. ricinus* (Überetsch) erbracht werden.

Rickettsien

Sporadisch klinische Fälle, serologischer Nachweis bei risikoexponierten Personen:

In einer Gesamttiroler Studie, INTERREG IV Projekt - VEIT Vektorassoziierte Erkrankungen Inneralpiner Täler – VEIT, wurden Rickettsien bei berufsexponierten Personen AK nachgewiesen: *Rickettsia helvetica* u. *Rickettsia* s.n.

Kultureller Nachweis aus *I. ricinus*: *R. sibirica* u. *R. conori*; aus *Haemaphysalis punctata*:

R. conori

NOTES / NOTIZEN

PARASITOLOGICAL DAY 2015

List of participants (attended) in alphabetic order

Name	Address	E-Mail
BUTTER Julia	Institut für Parasitologie Department für Pathobiologie Veterinärmedizinische Universität Wien Veterinärplatz 1, A-1210 Wien	julia.butter@icloud.com
DUSCHER Georg	Institut für Parasitologie Department für Pathobiologie Veterinärmedizinische Universität Wien Veterinärplatz 1, A-1210 Wien	georg.duscher@vetmeduni.ac.at
FÜHRER Hans-Peter	Institut für Parasitologie Department für Pathobiologie Veterinärmedizinische Universität Wien Veterinärplatz 1, A-1210 Wien	hans-peter.fuehrer@vetmeduni.ac.at
GLAWISCHNIG Walter	Austrian Agency for Health and Food Safety Technikerstrasse 70, A-6020 Innsbruck	walter.glawischnig@ages.at
HODZIC Adnan	Institut für Parasitologie Department für Pathobiologie Veterinärmedizinische Universität Wien Veterinärplatz 1, A-1210 Wien	adnan.hodzic@vetmeduni.ac.at
HÖRWEG Christoph	Naturhistorisches Museum Wien 3. Zoologische Abteilung Burgring 7, A-1010 Wien	christoph.hoerweg@nhm-wien.ac.at
HOHENSEE Nadine	Institute of Specific Prophylaxis and Tropical Medicine Medical University Vienna Kinderspitalgasse 15, A-1090 Wien	nadine@hohensee.de
HUSCH Christian	Institute of Specific Prophylaxis and Tropical Medicine Medical University Vienna Kinderspitalgasse 15, A-1090 Wien	a0805354@unet.univie.ac.at
KOTHMAYER Michael	Institut für Parasitologie und Zoologie Department für Pathobiologie Veterinärmedizinische Universität Wien Veterinärplatz 1, A-1210 Wien	michi.kothmayer@gmx.at
LEITNER Natascha	Institut für Parasitologie Department für Pathobiologie Veterinärmedizinische Universität Wien Veterinärplatz 1, A-1210 Wien	l.leitner@aon.at
MANSFELD Michael Dieter	ILV Kärnten, Veterinärmedizinische Untersuchungen Kirchengasse 43, A-9020 Klagenfurt	dieter.mansfeld@ktn.gv.at
MROWIETZ Naïke	Institut für Parasitologie Department für Pathobiologie Veterinärmedizinische Universität Wien Veterinärplatz 1, A-1210 Wien	1145142@students.vetmeduni.ac.at
SCHWARZMANN Laurin	Institut für Parasitologie Department für Pathobiologie Veterinärmedizinische Universität Wien Veterinärplatz 1, A-1210 Wien	laurin_schwarzmann@hotmail.com

PARASITOLOGICAL DAY 2015

Name	Address	E-Mail
SIMEONI Josef	Dienst f. Hygiene u. öffentl. Gesundheit des Gesundheitsbezirkes Bozen Amba Alagistr. 33, I-39100 Bozen	josef.simeoni@sabes.it
STENICO Alberta	Umweltagentur Autonome Provinz Bozen Unterbergstr. 2, I-39055 Leifers (Bozen)	alberta.stenico@provinz.bz.it
WALDER Gernot	Labor für Infektionsdiagnostik Dr. Gernot Walder GmbH A-9931 Außervillgraten 30	gernot.walder@i-med.ac.at
WEILER Martin	ABC-Abwehrschule ÖBH Platz der Eisenbahnpioniere 1, A-2100 Korneuburg	martin.weiler@gmx.at
ZIMMANN Felix	Universität Wien	f.zimm@upag.eu

Thanks for the support to



LABOR FÜR VETERINÄRMEDIZINISCHE DIAGNOSTIK UND HYGIENE GmbH

A-1030 WIEN RENNWEG 95
TEL.: (01)799 62 29 FAX: (01)799 62 29-50



Gemeinde Außervillgraten



naturhistorisches museum wien



Verein für
veterinärmedizinische
Parasitologie

SAVE THE DATE

**49. Jahrestagung der Österreichischen Gesellschaft für
Tropenmedizin, Parasitologie und Migrationsmedizin
(ÖGTPM)**

***49th Annual Meeting of the Austrian Society of Tropical Medicine,
Parasitology and Migration Medicine (ASTPM)***

Breakthroughs

**12. – 14. November 2015
Naturhistorisches Museum Wien
www.ogtpm.at**

